

## SESQUITERPENOID STRESS COMPOUNDS FROM *NICOTIANA* SPECIES

REIKO UEGAKI, TAKANE FUJIMORI, SUSUMU KUBO and KUNIO KATO

Central Research Institute, The Japan Tobacco and Salt Public Corporation,  
6-2 Umegaoka, Yokohama, 227, Japan

(Received 31 July 1980)

**Key Word Index**—*Nicotiana*; Solanaceae; stress compounds; solavetivone; 3-hydroxysolavetivone; solanascone; phytuberin; phytuberol; glutinosone; tobacco mosaic virus; tobacco rattle virus.

**Abstract**—Solavetivone, 3-hydroxysolavetivone, solanascone, phytuberin and phytuberol were identified as stress compounds in leaves of *Nicotiana tabacum* cv Samsun NN, *N. sylvestris*, which is the maternal progenitor of *N. tabacum*, produced all the above compounds except 3-hydroxysolavetivone. In the  $F_1$  hybrid of *N. tabacum* and *N. glutinosa*, all the stress compounds produced by *N. tabacum* and *N. glutinosa*, respectively, were accumulated.

### INTRODUCTION

The Solanaceae has been known to produce many structurally-related sesquiterpenoid stress compounds [1, 2]. To date, five sesquiterpenoid stress compounds, capsidiol [3], solavetivone [4, 5], phytuberin [6, 7], phytuberol [7, 8] and glutinosone [9], have been isolated from plants of the genus *Nicotiana*. In response to virus, only solavetivone accumulates in leaves of *Nicotiana tabacum* [4, 5], glutinosone in *N. glutinosa* [9] and capsidiol in *N. clevelandii* and *N. tabacum* [3].

In this paper, we report the occurrence of stress compounds in leaves of *N. tabacum* cv Samsun NN inoculated with tobacco mosaic virus (TMV), *N. sylvestris* inoculated with TMV and tobacco rattle virus (TRV) and the  $F_1$  hybrid of *N. tabacum* and *N. glutinosa* inoculated with TMV.

### RESULTS AND DISCUSSION

#### *N. tabacum* cv Samsun NN inoculated with TMV

The steam-distillate of the  $\text{CH}_2\text{Cl}_2$  extract from TMV-infected leaves contained solavetivone (1), 3-hydroxysolavetivone (3-hydroxyspirovetiva-1(10),11-dien-2-one) (2), solanascone (3), phytuberin (4) and phytuberol (5), which was revealed by capillary GC/MS analysis. These compounds were isolated as oils by preparative GLC. The mass,  $^1\text{H}$  NMR and IR spectra of 3-hydroxysolavetivone (2) and solanascone (3) were in accord with those of the reported data [10, 11]. The mass and IR spectra and retention times of the remaining compounds agreed with those of authentic samples. TMV stimulated maximum accumulations of 1.11 mg of solavetivone (1) and 0.68 mg 3-hydroxysolavetivone (2)/g dry wt of inoculated leaves during 7 days after inoculation. Concentrations of other compounds were as follows: solanascone (3) 0.06 mg, phytuberin (4) 0.14 mg and phytuberol (5) 0.21 mg/g dry wt of inoculated leaves.

#### *N. sylvestris* inoculated with TMV and TRV

Both steam distillates of the  $\text{CH}_2\text{Cl}_2$  extracts from leaves inoculated with TMV and TRV contained solavetivone (1), solanascone (3), phytuberin (4) and

phytuberol (5). The presence of these compounds was suggested by capillary GC/MS analysis and these compounds were identified by the mass spectra and retention times. 1 was present as the major metabolite in these inoculated leaves. Up to 1.52 (1.86\*) mg 1/g dry wt of TMV inoculated leaves and 1.02 (1.25\*) mg/g dry wt of TRV inoculated leaves were found. The accumulations of 3–5 were very low compared with that of 1. Thus, *N. sylvestris*, which is the maternal progenitor of *N. tabacum*, produced all the stress compounds of *N. tabacum* except 2.

#### *N. tabacum* × *N. glutinosa* inoculated with TMV

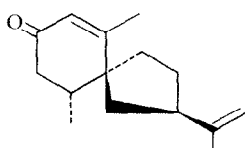
The presence of 1–6 in the steam distillate of the  $\text{CH}_2\text{Cl}_2$  extract from TMV-infected leaves were recognized by capillary GC/MS analysis. 1, 2 and 6 were the major stress compounds in inoculated leaves. Up to 0.13 mg 1, 0.05 mg 2 and 0.58 mg 6/g dry wt of inoculated leaves were found. The amounts of 3–5 were very low compared with the above three compounds. 6 in leaves of *N. glutinosa* inoculated with TMV was found at the level of 0.47 mg/g dry wt of inoculated leaves. Thus, the  $F_1$  hybrid of *N. tabacum* and *N. glutinosa* appeared to accumulate all the stress compounds produced by *N. tabacum* and *N. glutinosa* [9], respectively.

#### Uninoculated plants

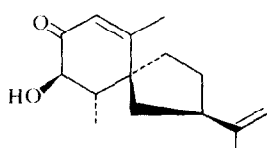
Sesquiterpenoid stress compounds were not detected by capillary GC/MS analysis in healthy leaves of *N. tabacum* cv Samsun NN, *N. sylvestris* and the  $F_1$  hybrid of *N. tabacum* and *N. glutinosa*.

Of these six compounds, 3-hydroxysolavetivone (2) and solanascone (3) have not been reported before as stress compounds in tobacco plants. 3-Hydroxysolavetivone (2) has been isolated as the aglycone of glucoside from flue-cured Virginia tobacco [10] and solanascone (3) has been obtained from air-cured Burley tobacco [11] and *N. sylvestris* [12] in very small amount.

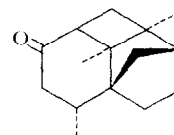
\* 1/g dry wt of leaves excluding the midrib.



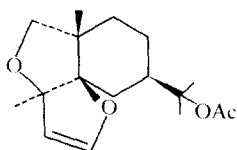
1 Solavetivone



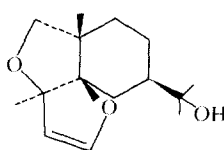
2 3-Hydroxysolavetivone



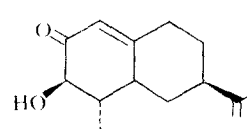
3 Solanascone



4 Phytuberin



5 Phytuberol



6 Glutinosone

### EXPERIMENTAL

*N. tabacum* cv Samsun NN was grown in a greenhouse at 24°. Fully expanded leaves of 2-month-old tobacco were inoculated with TMV-OM (0.5 µg/ml in 0.1 M phosphate buffer, pH 7.0). Carborundum was used as an abrasive. When brown lesions had been produced (7 days), the leaves were harvested. The harvested leaves (320 g fr. wt) which had been inoculated with TMV were freeze-dried. The dried materials (37 g) were extracted with CH<sub>2</sub>Cl<sub>2</sub> (21 × 3) and evapd to dryness. The crude extract (2.20 g) was steam-distilled and extracted with Et<sub>2</sub>O by simultaneous distillation and extraction [13]. The ether extract was evapd to dryness to give 218 mg of the volatile.

Similarly, the leaves of *N. sylvestris* (2-month-old), inoculated with a tomato strain of TMV (10 µg/ml in 0.1 M phosphate buffer, pH 7.0) or with the buffer extract (1:20) of TRV-infected tobacco leaves, were harvested. The harvested leaves were 550 g (TMV-infected) and 160 g (TRV-infected). The dried materials (TMV-infected 50 g and TRV-infected 27 g), crude extract (3.65 and 1.58 g) and volatile (158 and 89 mg) were obtained.

Tobacco leaves of *N. tabacum* × *N. glutinosa* (2-month-old) were inoculated with TMV (0.3 µg/ml in 0.1 M phosphate buffer, pH 7.0). Six days later, the leaves were harvested. The harvested leaves (210 g fr. wt) were treated in the same manner described above. The dried materials (26 g), crude extract (1.60 g) and volatile (91 mg) were obtained.

The leaves of 2-month-old *N. glutinosa* were inoculated with TMV (0.5 µg/ml in 0.5 M phosphate buffer). The fresh leaves (600 g), dried materials (70.5 g), CH<sub>2</sub>Cl<sub>2</sub> extract (4.4 g) and steam distillate (151 mg) were obtained.

1–6 were isolated as an oil from the volatile of virus-inoculated tobacco leaves by prep. GLC (5% FFAP 3 mm × 1 m, 100–240°, 5°/min). The following spectral data of 2 and 3 were obtained. 3-Hydroxysolavetivone (2): MS *m/z* (rel. int.): 234 (M<sup>+</sup>, 5), 205 (9), 176 (100), 161 (32), 133 (49), 109 (70), 108 (70), 91 (53), 68 (88); <sup>1</sup>H NMR (100 MHz, CDCl<sub>3</sub>): δ 1.21 (3 H, *d*, *J* = 7 Hz), 1.75 (3 H, *s*), 2.02 (3 H, *s*), 3.80 (1 H, *d*, *J* = 12.5 Hz), 4.75 (2 H, *s*), 5.81 (1 H, *m*); IR  $\nu_{\max}^{\text{film}}$  cm<sup>-1</sup>: 3480, 3080, 1680, 1645, 890. Solanascone (3): MS *m/z* (rel. int.): 218 (M<sup>+</sup>, 28), 203 (10), 190 (62), 121 (49), 120 (100), 105 (37), 91 (31), 41 (45); <sup>1</sup>H NMR (100 MHz, CDCl<sub>3</sub>): δ

1.03 (3 H, *s*), 1.11 (3 H, *d*, *J* = 7 Hz), 1.31 (3 H, *s*), 2.43 (2 H, *m*); IR  $\nu_{\max}^{\text{film}}$  cm<sup>-1</sup>: 1710, 1460, 1426, 1382, 1300.

Characteristic retention times on capillary GLC (PEG-20M Ultra Bond 0.27 mm × 60 m, 100–215°, 2°/min) were 1 43.6 min, 2 51.6 min, 3 33.2 min, 4 40.0 min, 5 43.1 min and 6 47.7 min. The amounts of 1 and 6 were estimated by the mass fragmentography procedure and those of the other compounds were evaluated by integration of the peaks in the capillary gas chromatograms.

### REFERENCES

1. Stoessl, A., Stothers, J. B. and Ward, E. W. B. (1976) *Phytochemistry* **15**, 855.
2. Masamune, T., Murai, A. and Katsui, N. (1979) *Kagaku To Seibutsu Jpn.* **16**, 648.
3. Bailey, J. A., Burden, R. S. and Vincent, G. G. (1975) *Phytochemistry* **14**, 597.
4. Ito, T., Takahashi, T., Oshima, Y., Takusari, H. and Odagiri, S. (1979) *Agric. Biol. Chem.* **43**, 413.
5. Fujimori, T., Uegaki, R., Takagi, Y., Kubo, S. and Kato, K. (1979) *Phytochemistry* **18**, 2032.
6. Hammerschmidt, R. and Kuč, J. (1979) *Phytochemistry* **18**, 874.
7. Uegaki, R., Fujimori, T., Kaneko, H., Kubo, S. and Kato, K. (1980) *Phytochemistry* **19**, 1543.
8. Uegaki, R., Fujimori, T., Kaneko, H., Kubo, S. and Kato, K. (1980) *Phytochemistry* **19**, 1229.
9. Burden, R. S., Bailey, J. A. and Vincent, G. G. (1975) *Phytochemistry* **14**, 221.
10. Anderson, R. C., Gunn, D. M., Murray-Rust, J., Murray-Rust, P. and Roberts, J. S. (1977) *J. Chem. Soc. Chem. Commun.* 27.
11. Fujimori, T., Kasagu, R., Kaneko, H., Sakamura, S., Noguchi, M., Furusaki, A., Hashiba, N. and Matsumoto, T. (1978) *J. Chem. Soc. Chem. Commun.* 563.
12. Wallin, I., Narbonne, C., Wahlberg, I., Nishida, T. and Enzell, C. R. (1980) *Acta Chem. Scand.* **34**, 391.
13. Schultz, T. H., Flath, R. A., Mon, T. R., Eggling, S. B. and Teranishi, R. (1977) *J. Agric. Food Chem.* **25**, 446.